General and Comparative Endocrinology 265 (2018) 214-218



Contents lists available at ScienceDirect

General and Comparative Endocrinology

journal homepage: www.elsevier.com/locate/ygcen

Review article

Dual function model revised by thyroid hormone receptor alpha knockout frogs



^a Department of Biological Sciences, University of Cincinnati, Cincinnati, OH, USA

^b Molecular Morphogenesis, National Institute of Child Health and Human Development, National Institutes of Health, Bethesda, MD, USA

ARTICLE INFO

Keywords: Xenopus tropicalis Tadpole Metamorphosis Gene knockout

ABSTRACT

All vertebrates require thyroid hormone (TH) for normal growth and development. Plasma TH enters cells and alters gene expression via nuclear receptors TR α and TR β . *In-vitro* studies showed that TRs function as repressors of TH-inducible genes in the absence of TH and as activators of those same genes in the presence of TH. A dual function model was proposed to harmonize these molecular TR actions with the dynamic expression of TRs and peak in production of TH experienced during development. Conclusive tests of the repression activity of TRs early in development as predicted by the model awaited gene knockout technology targeting TR α . At the molecular level, active repression of genes involved in metamorphosis by TR α in the absence of TH was confirmed in whole bodies and intestine from TR α knockout studies. As a consequence of this reduced repression in TR α knockout animals, initiation of limb morphogenesis occurs precociously. However, subsequent limb development is retarded during rising plasma TH levels due to reduced TR-dependent responsivity to TH. In contrast to the limbs, intestine remodeling is delayed by one to two developmental stages in TR α knockout animals, despite de-repressed levels of TH-induced genes during premetamorphosis. Surprisingly, in the absence of TR α , hind limbs do not require gene induction by TH signaling to complete morphological growth and development, which is contrary to prediction by the dual function model. Full evaluation of the dual function model for all organs awaits the production of TR α and TR β double knockout frogs.

1. Introduction

Thyroid hormone (TH) plays critical roles in all vertebrates during development and adult physiology (Braverman and Utiger, 2005; Yen, 2001). All vertebrates experience a peak in plasma levels of TH, often at a life history transition, such as birth in humans, weaning in mice, and metamorphosis in amphibians (Buchholz et al., 2006; Laudet, 2011). Lack of TH during development leads to severe growth and neurological impairments in humans, death around weaning in mice, and complete lack of metamorphosis in frogs. The actions of TH are mediated by TRs, which are members of the nuclear hormone receptor superfamily (Cheng et al., 2010). All vertebrates have two TR isoforms: TRa and TR β (Laudet, 2011). TR α is more wide-spread among tissues and is typically expressed before thyroid gland development when TH is minimally available to tissues (Cheng et al., 2010; Forhead and Fowden, 2014; Ng and Forrest, 2006). TRs act by binding TREs in the enhancers of TH response genes to regulate their transcription. For THinduced genes, TRs heterodimerize with retinoid X receptors (RXRs) to recruit corepressors in the absence of TH to actively repress genes, and TR / RXR heterodimers recruit coactivators in the presence of TH to induce gene expression (Buchholz et al., 2006; Cheng et al., 2010; Shi et al., 2012).

Based on these molecular actions of TR derived mainly from in-vitro cell culture studies, a dual function model to describe the role of TR in development during pre- vs. metamorphosis was first proposed upon cloning of the frog TRs and analyzing their developmental expression (Yaoita and Brown, 1990). As elaborated in subsequent reviews (Buchholz et al., 2006; Sachs et al., 2000; Shi et al., 1996; Shi, 2009), the dual function model states that unliganded TR represses genes during premetamorphosis when TH is virtually absent to allow larval growth and that TR induces those same genes when TH is present to bind TR to enable TH-dependent metamorphosis. Given the low levels of TH during early stages of metamorphosis and the different levels of sensitivity to TH among tissues (Choi et al., 2015a; Leloup and Buscaglia, 1977; Shi et al., 1996), the transition from unliganded to liganded TR is not simultaneous among tissues but is achieved at different developmental time points. For instance, it is likely that at the beginning of metamorphosis TR is an activator in limbs while still a repressor in tail due to different levels of free

^{*} Corresponding authors.

E-mail addresses: buchhodr@ucmail.uc.edu (D.R. Buchholz), shi@helix.nih.gov (Y.-B. Shi).

https://doi.org/10.1016/j.ygcen.2018.04.020

Received 29 September 2017; Received in revised form 14 April 2018; Accepted 14 April 2018 Available online 22 April 2018 0016-6480/ © 2018 Elsevier Inc. All rights reserved.

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intracellular TH.

The dual function model also addresses the developmental consequences of the molecular actions of TR, where TR-mediated gene repression allows premetamorphic animals to grow and prevents precocious metamorphosis until plasma TH becomes available to induce TH-response genes and stimulate development to proceed through metamorphosis. Numerous experiments in wild-type and transgenic animals supported the dual function model (Shi, 2013). However, the evidence for TR-mediated repression remained equivocal and the requirement for induced levels of TH-response gene expression for developmental progression remained untested (Choi et al., 2015b). Direct evaluation of the dual function model using TR gene knockout became possible with the advent of TALEN and CRISPR gene disruption technology in frogs (Blitz et al., 2013; Guo et al., 2014; Ishibashi et al., 2012; Lei et al., 2012; Nakayama et al., 2013; Suzuki et al., 2013).

TR α knockout (TR α KO) animals were analyzed in two papers by Choi et al. (2015b, 2017), two papers by Wen and Shi (2015), Wen et al. (2017), a review by Wen and Shi (2016), and 3 News & Views (Sachs, 2015; Schreiber, 2017; Yen, 2015). In both laboratories, the TR α DNA binding domain was targeted using TALENs. Frameshift mutations at this location were produced and expected to result in a truncated protein with no known activity. In all papers, overall growth and development as well as gene expression in whole body or tissues were analyzed, but various methods and experimental aims were present in each paper (Table 1).

2. Molecular level analysis of TRaKO

Expectations for altered regulation of gene expression in TRaKO tadpoles were based on the knowledge that before metamorphosis TRa is the predominant TR expressed in cells (Baker and Tata, 1990; Eliceiri and Brown, 1994; Yaoita and Brown, 1990). TR β has low expression at this time due to an alternate promoter (Shi et al., 1992), but most TR β expression is due to it being a direct TH response gene (Kanamori and Brown, 1992; Machuca et al., 1995; Ranjan et al., 1994). Consistent with premetamorphic expression of TR binding to TH response elements (TREs) both in the absence and presence of ligand, it was found that in premetamorphic TR α KO tadpoles, ChIP analysis using anti-TR and anti-NCoR (nuclear corepressor) antibodies showed a large reduction in TR binding to TR β and TH/bZIP TREs associated with reduced NCoR recruitment, as was expected due to lack of wild-type TRa (Wen et al., 2017). Residual significant binding by TR and NCoR to TREs above background levels in TRaKO animals is likely due to wild-type $TR\beta$ which is also recognized by the anti-TR antibodies and recruits NCoR (Sachs and Shi, 2000; Sachs et al., 2002). After 18 hrs. of TH treatment which induces $TR\beta$ expression by greater than 10 fold in wild-type animals, recruitment of TRs is increased in wild-type and TRaKO tadpoles (Wen et al., 2017), but the degree of increase is less in TRaKO compared to wild-type tadpoles, likely due to reduced level of TRE occupancy in the absence of TRa.

Expected changes in gene expression due to reduced TRE occupancy and coregulator recruitment in $TR\alpha KO$ tadpoles were reduced

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repression in premetamorphosis as well as reduced activation of THresponse genes when given exogenous TH. Indeed, higher expression levels, i.e., de-repressed expression levels, of TRB, TH-induced bZIP protein (TH/bZIP), Krüppel-like factor 9 (klf9), and stromelysin 3 (ST3) were observed in whole bodies of TRaKO premetamorphic animals (Choi et al., 2015b; Wen and Shi, 2015; Wen et al., 2017). Similarly, higher expression levels of the direct response genes TRB, KLF9, ST3, and sonic hedgehog (xhh) were observed in intestines isolated from untreated premetamorphic TRaKO tadpoles (Choi et al., 2017). In response to exogenous TH, gene induction is expected to be reduced in TRaKO animals because of reduced overall TR binding to TREs for recruiting coactivators and inducing transcription. For whole bodies and intestines in premetamorphic animals, the degree of induction by exogenous TH of TRB, TH/bZIP, klf9, ST3, and xhh was reduced in TRaKO tadpoles (Choi et al., 2015b, 2017; Wen and Shi, 2015; Wen et al., 2017). The reduced but still significant induction in TRa mutants is likely due to low levels of TR β expression and autoregulation of TR β available to induce TH-response genes. Non-genomic actions of TH may also contribute to TH-response gene induction (Cheng et al., 2010).

Significant differences in TH response gene expression were found throughout larval development in intestines in TR α KO compared to wild-type tadpoles (Choi et al., 2017). Gene induction for TR β and KLF9 was delayed by at least three developmental stages, and maximal induction was significantly reduced for ST3. For xhh, its expression was not maintained for as many developmental stages in TR α KO tadpoles. Each gene had a unique alteration of its expression profile, likely due to gene-specific regulators of expression, but in general there was delayed initiation and/or reduced and broadened shape of the expression profile across larval intestine development and metamorphosis.

3. Growth and development in TRaKO animals

The significant effects on gene expression observed in TR α KO animals were expected to have significant effects on phenotype. Studies so far using TR α KO animals have examined hind limb, intestine, overall growth, and timing of developmental events (Choi et al., 2015b, 2017; Wen and Shi, 2015; Wen et al., 2017). A surprising finding, given the high TR α gene expression levels throughout the tadpole across developmental stages, was that TR α KO animals can achieve complete metamorphosis and become fertile adults (Choi et al., 2017; Wen et al., 2017). Significant TR α KO growth and development phenotypes during the metamorphic process have been detected, but they have so far been seemingly mild and transitory, reminiscent of phenotypes seen in TR α mutant mice (Gauthier et al., 2001).

In general, the growth trajectories are very similar between wild-type and TR α KO animals (Fig. 1A) (Choi et al., 2017). However, during the first 2–3 weeks after fertilization, TR α KO animals are significantly larger than wild type (Choi et al., 2017; Wen et al., 2017). This growth difference is correlated with a difference in growth hormone mRNA expression level in 11 day old tadpoles, where the larger TR α KO tadpoles express more growth hormone as measured in whole bodies (Wen et al., 2017). Such an effect in TR α KO animals may be due to the role of TR α on growth hormone expression known from mammalian studies where growth hormone is a direct response gene, and lack of repression due to mutant TR α may

 Table 1 Comparison of Methods and Experimental Aims among TRaKO articles.

Article	Genotyping Method	Source	Developmental Stage/Organs
Choi et al. (2015a,b)*	limb phenotype	F1	premetamorphosis/limb, gills
Choi et al. (2017)*	limb phenotype	F1, F2	metamorphosis/intestine
Wen and Shi (2015)	PCR typing	founders	pre- and prometamorphosis/ limb
Wen et al. (2017)	PCR typing	F2 line	metamorphosis, limb, tail, intestine

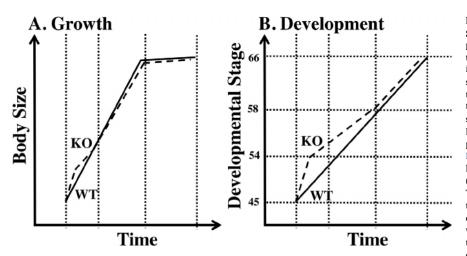
* Methimazole was used in Choi et al. (2015a,b, 2017).

allow higher expression of growth hormone in the absence of TH. During the remainder of the 6–7 week larval period through metamorphosis, snout-vent length was not statistically significantly different between TR α KO and wild-type tadpoles (Choi et al., 2017) but body weight was significantly reduced by about 10% at tail resorption at the end of metamorphosis in TR α KO animals (Wen et al., 2017).

One of the most significant findings using TR α KO animals was the effect on hind limb development (Fig. 2) (Choi et al., 2015b; Wen and Shi, 2015; Wen et al., 2017). The dual function model predicted that lack of repression by TRs may allow increased expression levels of TH response genes important for metamorphosis thereby allowing precocious development to occur. Indeed, hind limb morphogenesis

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Fig. 1. Diagrammatic representation of overall growth and development throughout the larval period and metamorphosis in wild-type and TRaKO tadpoles. A) TRaKO tadpoles (KO, dashed lines) initially grow faster than wild-type tadpoles (WT, solid lines) during premetamorphosis, but during the remainder of the larval period from the initiation of metamorphosis through to tail resorption, significant but small size differences are observed. B) TRaKO animals initially develop faster during premetamorphosis (< stage 54) (Nieuwkoop and Faber, 1994) than wild-type tadpoles based on hind limb criteria. Then, during prometamorphosis (stages 54-58) when TH levels begin to increase, TRaKO animals progress more slowly than wildtype tadpoles based on hind limb criteria. Then, during climax of metamorphosis (stages 58-66), when TH levels reach their peak, TRaKO and wildtype tadpoles develop at a similar rate. The vertical dotted lines from left to right in both graphs in-

dicate initiation of feeding (stage 45), initiation of metamorphosis (stage 54), mid-point of plasma TH levels during metamorphosis (stage 58), and tail resorption (stage 66).

initiated much earlier in TR α KO tadpoles compared to their wild-type siblings. This finding is the clearest demonstration of a developmental role for unliganded TR in vertebrate development and is consistent with the dual function model.

The limbs may be unique in the predominance of TRa compared to TR β , where high levels of TR α are expressed throughout the limb bud and TRβ expression is limited in hind limbs to cartilage based on *in-situ* studies (Cai and Brown, 2004; Fairclough and Tata, 1997; Shi et al., 1996). Other organs are likely less affected by TRaKO because of their higher expression of TR β which may in part or completely compensate for the loss of TRa (Wen et al., 2017). Thus, even though TRaKO animals initiate limb morphogenesis earlier in time (Fig. 1B), the other organs of the TR α KO tadpole may be in a state of developmental progression identical to wildtype tadpoles of the same age. On the other hand, in prometamorphosis (Nieuwkoop and Faber stages 54-58) (Nieuwkoop and Faber, 1994), there is a slow-down in developmental progression in TRaKO animals (Fig. 1B) (Choi et al., 2017; Wen et al., 2017). These stages are determined by the progress of limb morphogenesis, which may proceed more slowly due to the reduced rate of limb elongation from reduced responsivity to TH. Indeed, reduced limb elongation in response to exogenous TH treatment in TRaKO tadpoles is consistent with this possibility (Choi et al., 2015b). Later, during climax of metamorphosis (Nieuwkoop and Faber stages 58-66), the similarity in duration to progress through these stages in TRaKO and wild-type animals when plasma TH is highest may be due to

the TH signal (Fig. 1B).

Despite the effect of TR α KO on premetamorphic development (before stage 54) and prometamorphosis (stages 54–58), the total time from fertilization to the completion of metamorphosis (stages 1–66) in the TR α KO and WT animals was either the same (Wen et al., 2017) or slightly shorter (6 days out of 75 days) (Choi et al., 2017) (Fig. 1B). The somewhat different results may be explained by different methods and samples sizes. If TR α KO animals do metamorphose earlier, the mechanism is not clear and suggests that the opposing effects of TR α KO to initiate premetamorphosis earlier and delay progress through prometamorphosis fail to compensate for each other. Other possibilities include that perhaps TR α KO has the effect of increasing TH plasma levels, or derepression of TR β prior to initiation of metamorphosis resulted in increased development rate in the presence of TH.

The effect of TR α KO on intestinal remodeling reveals a different picture compared to the effect on hind limbs. The initiation of intestinal shrinkage occurred at a later developmental stage in TR α KO vs. wild-type animals and did not progress to the same extent during natural or TH-induced metamorphosis (Choi et al., 2017; Wen et al., 2017). Importantly, despite derepression of TH response genes in premetamorphic TR α KO intestines, intestinal remodeling was delayed rather than being precocious as in hind limb (Choi et al., 2017). Specifically, larval epithelial cell apoptosis was delayed by two stages, and the number of intestinal folds into the lumen was significantly

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compensation by sufficient induction of TRp in other organs to mediate

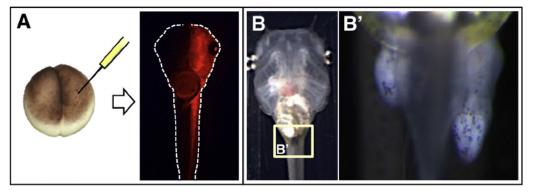
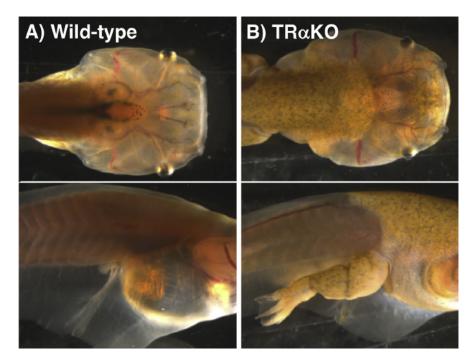


Fig. 2. Mutations in TRα result in precocious initiation of hind limb morphogenesis. A) Injection of TALEN mRNAs targeting the DNA-binding domain of TRα was performed in one cell of a two-cell stage embryo of *Xenopus tropicalis*. mCherry mRNA was co-injected as a red fluorescent tracer to identify which side of the embryo (left in this case) was injected with TRα TALENS. B) Hind limbs of the resulting tadpole during premetamorphosis were examined for developmental progress on the injected and uninjected sides of the animal. As shown in B', the hind limbs on the injected side are more advanced in size and stage, due to lack of TRα-mediated repression of genes important for metamorphic progression.

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Fig. 3. Hind limb and skin morphogenesis in the absence of TH signaling in TRaKO tadpoles. Wildtype (A) and TRaKO (B) tadpoles shown in dorsal and side views were reared in 1 µM methimazole (inhibits TH synthesis) for 10 weeks starting at the initiation of feeding when the thyroid gland is forming. Development in the methimazole-treated wild-type tadpole is blocked at the beginning of metamorphosis, at the point when plasma TH levels begin to rise in untreated tadpoles. Notably, hind limb morphogenesis has stalled just after the paddle stage, and the skin is still thin and transparent as it is before it remodels to adult skin. In the methimazoletreated TRaKO tadpole, hind limb morphogenesis is complete, and in locations where adult skin replaces larval skin (i.e., not skin covering gills and tail), the skin has become opaque obscuring view of internal structures

proliferation or cell size or intestinal diameter were detected. These intestinal phenotypes may reflect a permanent deficit in intestinal remodeling which may explain the slower growth observed after 4 weeks post-metamorphosis (Choi et al., 2017). However, as in TR α KO mice, the effect on growth and development and intestinal morphology appears to be minimal (Gauthier et al., 2001).

A surprising finding in the TR α KO animals was their morphology after prolonged treatment with methimazole, which blocks TH synthesis (Fig. 3). Continuous treatment with methimazole at the beginning of feeding when the thyroid gland is being formed can virtually eliminate the production TH in treated tadpoles (Buchholz et al., 2004; role of TH signaling is not to determine cell fate during development but rather to permit development that is already specified. Thus, a major action of TRs in development appears to be to control the timing of developmental initiation and rate of the change from the larval to adult type.

4. Conclusions

The dual function model was largely supported using TR α KO animals by studies reviewed here (Choi et al., 2015b, 2017; Wen and Shi, 2015;

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Buckbinder and Brown, 1993). Wild-type animals kept in methimazole stop developmental progression around stage 53–54, when the hind limbs are around the paddle stage. These animals continue to grow indefinitely in size, but they do not advance in stage, whereas their untreated siblings undergo metamorphosis after 5-7 weeks. TRaKO tadpoles treated with methimazole for over 10 weeks have a mixed phenotype (Choi et al., 2017). Specifically, when TRaKO tadpoles are reared in methimazole, the limbs, which express predominantly TRa and little TRB, experience a lack of TH signaling yet achieve complete morphogenesis (Fig. 3). This result indicates that derepressed levels, rather than induced levels, of response gene expression are sufficient to complete limb morphogenesis. The skin of methimazole-reared TRaKO tadpoles also appeared to complete development, except in skin covering gill and tail areas where adult skin does not form (Fig. 3) (Suzuki et al., 2009). Other parts of their external morphology remain larval, such as mouth parts, gills, and tail. Thus, in the complete absence of TH signaling, hind limbs and adult skin can form if TRa is absent, although the rate of limb and skin development may be affected. This result is surprising because according to the dual function model, TH response gene induction is required for full development to the adult form. Derepression is predicted to be most complete in hind limbs because they are dominated by TRa. The formation of adult skin in methimazoletreated TRaKO animals predicts that skin development is also predominantly dependent on TR α . The tail has a high proportion of TR β , and thus TR β in tail may be sufficient to maintain TH-response gene repression. Alternatively, tail may require TH response gene induction above levels of gene expression achieved by derepression. These fi ndings using methimazole further suggest that, at least for the limb, the

Wen et al., 2017). Molecular analyses from ChIP and gene expression supported functional gene repression by unliganded TR in early pre-metamorphosis. Reduced gene induction during metamorphosis and after exogenous TH treatment supported the role of gene activation by liganded TR. These effects on gene regulation are consistent with the dual function model and explain precocious initiation of hind limb development in TRaKO animals as well as delayed completion of limb morphogenesis. The relationship between the dual function model and initial faster growth but reduced weight at tail resorption is not as clear but may relate to TH regulation of pituitary hormones. The early intestine gene derepression followed by subsequent delay in intestine remodeling is also not explained but may be due to low levels of TRB to accomplish repression and reduced TRB autoregulation to reduce the rate of remodeling. The requirement for induced levels of TH-response gene expression for developmental progression as proposed by the dual function model was found to be not accurate for the limbs and in need of further evaluation in other organs. It appears that levels of TH-response gene expression equivalent to de-repressed levels when TR α is absent, rather than induced levels of these genes, are sufficient to achieve complete metamorphosis.

Our understanding of the TR α knockout phenotype would benefit from knowing when plasma TH becomes available in TR α KO animals and what level plasma TH achieves. However, early TH production does not explain precocious hind limb development, because TR α KO animals reared in methimazole, which blocks TH synthesis, exhibit the same precocious hind limb phenotype. Because hind limb development and nothing else seems to be precocious in TR α KO animals, it may not be appropriate to compare stage 54 wild-type animals to stage 54 TR α KO animals because the rest of the organs besides the limb in TR α KO stage 54 animals are not expected to be in an accelerated state of development.

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TR β is expressed at least to some degree in most if not all tissues, such that full evaluation of the dual function model will require TR α/β double knockout animals. It will be important to determine the degree to which other organs besides limb may exhibit precocious initiation of metamorphosis and be able to complete development in the complete absence of TR-mediated repression and activation. In this regard, it is interesting to note that TR α/β double knockout mice have reduced viability and significant morphological and physiological defects, even though mouse development is less dependent on TH than frog metamorphosis (Gothe et al., 1999). Given the complete limb morphogenesis without TH signaling and despite the extraordinary dependence of metamorphosis on TH, it is an open question whether completion of metamorphosis through tail resorption in the absence of TRs will occur.

Note added in proof

A recent article by Nakajima et al. (2018) reported results using TR α KO animals entirely consistent and supportive of the conclusions of the current review.

Acknowledgement

This work was supported in part by the Intramural Research Program of NICHD, NIH (YS).

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at https://doi.org/10.1016/j.ygcen.2018.04.020.

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